

Substituted 2-Azabicyclo[2.1.1]hexanes as Constrained Proline Analogues: Implications for Collagen Stability

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Received May 4, 2004

Among the proteinogenic amino acids, only proline is a secondary amine and only proline has a saturated ring. Electronegative substituents on C-4 (that is, C^γ) have a substantial effect on the trans/cis ratio of the prolyl peptide bond and the pucker of the pyrrolidine ring. 2-Azabicyclo[2.1.1]-hexane is, in essence, a proline analogue with two C^γ atoms, one in each of the two prevalent ring puckers of proline. Here, 2-azabicyclo[2.1.1]hexane analogues of 2S-proline, (2S,4S)-4-hydroxyproline, and (2S,4S)-4-fluoroproline residues were synthesized, and their trans/cis ratios were shown to be invariant in a particular solvent. Thus, the substitution of a proline residue on C-4 affects the trans/cis ratio by altering the pucker of its pyrrolidine ring. This finding has implications for the conformation of collagen, which has an abundance of 2S-proline and (2S,4R)-4-hydroxyproline residues, and can be stabilized by (2S,4R)-4-fluoroproline and (2S,4S)-4-fluoroproline residues.

Introduction

Proline has two prominent attributes that are unique among the proteinogenic amino acids: only proline is a secondary amine, and only proline has a saturated ring.¹ These attributes make proline residues a key determinant of protein structure.^{2,3} Accordingly, a deeper understanding of the conformational properties of proline would illuminate challenging problems in protein folding, stability, and design.

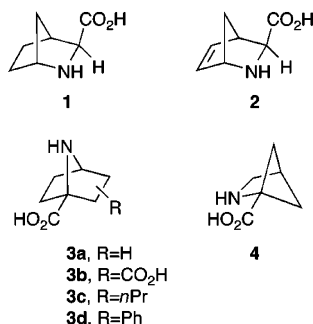
As a secondary amine, proline has a much greater propensity than other natural amino acids to form cis (that is, *E*) peptide bonds.^{4–6} A variety of methods have been developed to control the trans/cis ratio, including

buttressing the 2-,⁷ 3-,^{7,8} and 5-positions^{9–12} with functional groups, replacing the prolyl peptide bond with an alkene isostere,^{13–18} and including the amide in a ring system that is fused to the pyrrolidine ring.¹¹ These approaches endow torsional control of the amide bond but introduce steric bulk that could be undesirable.

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The pyrrolidine ring of proline exists in a variety of puckers, with C γ being its most aplanar constituent.^{19,20} A variety of bridged bicyclic proline mimics have been developed to control the conformation of the pyrrolidine ring (1–4).^{21–27} All of these proline mimics are rigid enough to fix pyrrolidine ring pucker, but some include elements that make them suboptimal as proline mimics. Of those proline mimics, **3a**^{28,29} and **4**³⁰ have been employed in the design of peptide-based enzyme inhibitors to reinforce a bioactive peptide conformation, with varying degrees of success.



2-Azabicyclo[2.1.1]hexane (as in **5**) is a proline analogue that displays *both* predominant puckers of the pyrrolidine ring.³¹ This end is achieved by the addition of a single carbon atom to proline, a minimal perturbation. Substitution of a hydrogen at the C γ^1 or C γ^2 position of the bicyclic system with a hydroxyl or fluoro group yields mimics of 4-hydroxyproline (as in **6**) and 4-fluoroproline (as in **7**). (2*S*,4*R*)-4-Hydroxyproline (Hyp) residues are prevalent in collagen, which is the most abundant protein in animals.³² Replacing the Hyp residues with (2*S*,4*R*)-4-fluoroproline (Flp) residues endows synthetic

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TABLE 1. Values of $K_{\text{trans/cis}}$ for 4-Substituted AcXaaOMe^{a,b}

Xaa	X	Y	$K_{\text{trans/cis}}$
Flp	H	F	6.7
Hyp	H	OH	6.1
Pro	H	H	4.6
hyp	OH	H	2.4
flp	F	H	2.5

^a Data are from ref 38. ^b Values were measured in D₂O at 25 °C by integration of ¹H NMR spectra.

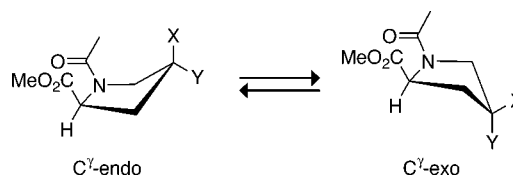
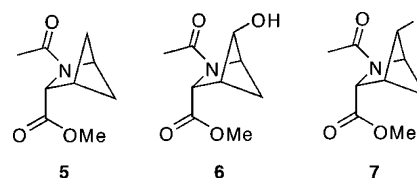


FIGURE 1. Ring puckers in 4-substituted Ac-Pro-OMe. C γ -endo pucker is favored when X = H, OH, or F and Y = H. C γ -exo pucker is favored when X = H and Y = OH or F.

mimics of collagen with extraordinary stability.^{33–35}



Substitutions on C-4 (that is, C γ) of proline residues are known to have a large effect on the trans/cis ratio.^{36–39} For example, electronegative substituents in the 4*R*-position of Pro increase the stability of the trans isomer, whereas electronegative substituents in the 4*S*-position decrease that stability (Table 1). NMR analyses indicate that Ac-(2*S*,4*R*)-4-fluoroproline-OMe (Ac-Flp-OMe) resides predominantly (86%) in the C γ -exo pucker in solution, whereas Ac-(2*S*,4*S*)-4-fluoroproline-OMe (Ac-flp-OMe) is found almost exclusively (95%) in the C γ -endo pucker (Figure 1).⁴⁰ This dichotomy can be attributed to the gauche effect, which causes the pyrrolidine ring to adopt a pucker that places the nitrogen and fluorine in a gauche orientation about the C δ^2 –C γ bond (for nomenclature, see Figure 2 and ref 41)⁴² and has important

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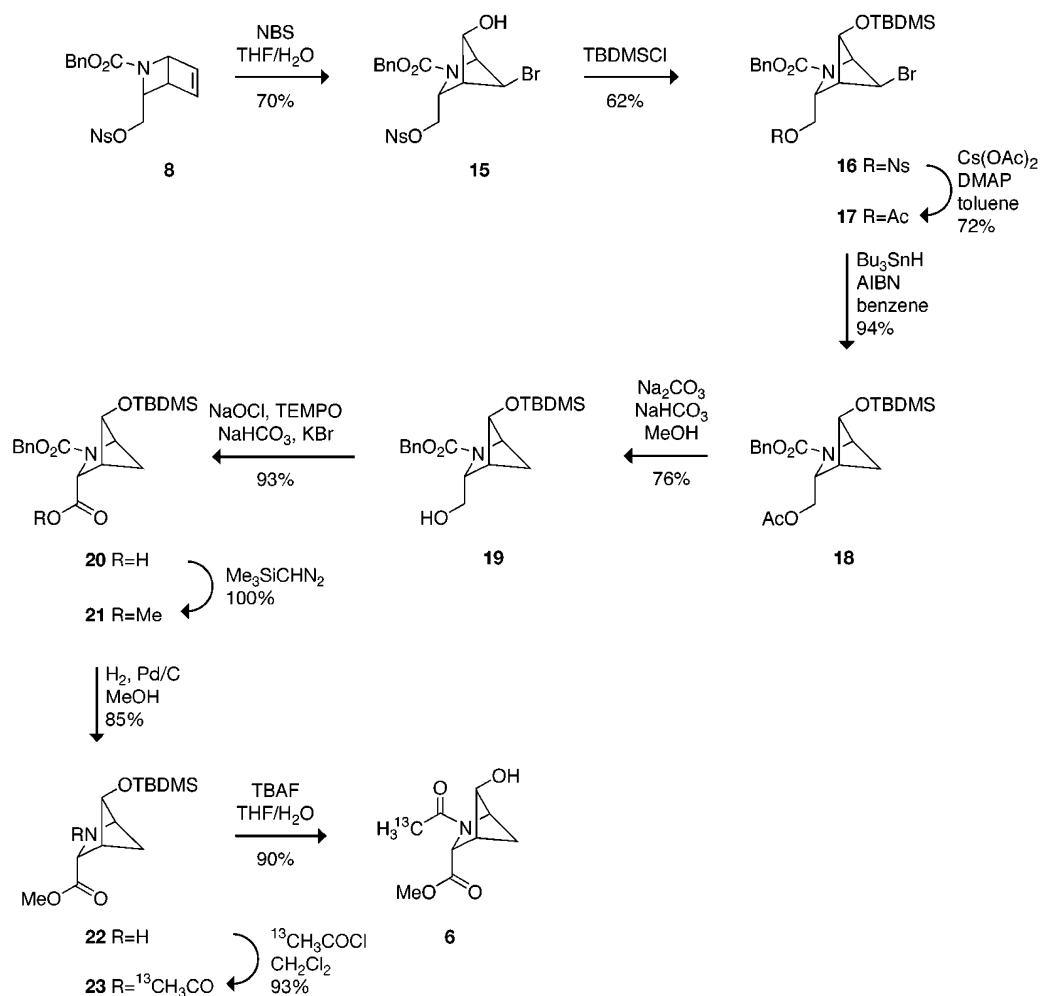
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SCHEME 2. Synthetic Route to Ac-methano-hyp-OMe (6)



hexane **9** along with a tricyclic byproduct, **10**, in a ratio of 2:1.^{48–50} To generate the hydroxyl group of alcohol **13** in high yield, the *p*-nitrobenzenesulfonyl (nosyl) group of **9** was replaced with an acetyl group by reaction with cesium acetate⁵¹ to yield **11**. The bromine groups were then removed by reduction with tributyltin hydride initiated by AIBN^{48,49} to give ester **12**. The acetate was hydrolyzed under mildly basic conditions to alcohol **13**, followed by oxidation of the alcohol to the carboxylic acid⁵² and esterification with trimethylsilyldiazomethane⁵³ to yield compound **14**. The benzyloxycarbonyl group was removed by hydrogenolysis under standard conditions and acetylated with ¹³C-labeled acetyl chloride to give the desired Ac-methano-Pro-OMe (**5**). The ¹³C label was incorporated to enable facile measurement of amide trans/cis ratios by ¹³C NMR spectroscopy (vide infra).

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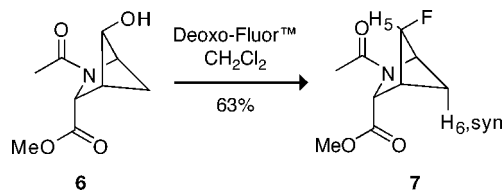
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SCHEME 3. Synthetic Route to Ac-methano-flp-OMe (7)



The synthesis of 2-(2-¹³C-acetyl)-5-hydroxy-2-azabicyclo[2.1.1]hexane-3-carboxylic acid methyl ester (**6**, Ac-methano-hyp-OMe) was similar to that of methano-Pro. The key differences include effecting the rearrangement from **8** to **15** with NBS and H₂O in THF⁵⁰ and the need to protect the resulting alcohol with a TBDMS group, as shown in Scheme 2. The relative configuration at C₁'¹ (which bears the hydroxyl group) of **6** was determined by X-ray diffraction analysis (vide infra).

2-Acetyl-5-fluoro-2-azabicyclo[2.1.1]hexane-3-carboxylic acid methyl ester (**7**, Ac-methano-flp-OMe) was synthesized from unlabeled Ac-methano-hyp-OMe (**6**) by its reaction with bis(2-methoxyethyl)aminosulfur trifluoride (Deoxo-Fluor), as shown in Scheme 3.⁵⁴ The reaction generally proceeds with the inversion of stereochemis-

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TABLE 2. Effect of Solvent on $K_{\text{trans/cis}}$ of AcXaaOMe^a

Xaa	$K_{\text{trans/cis}}$		
	CDCl ₃	1,4-dioxane- <i>d</i> ₈	D ₂ O
methano-Pro (5)	2.4	2.2	3.5
methano-hyp (6)	2.4	2.1	3.6
methano-flp (7)	2.7	2.8	3.5

^a Values of $K_{\text{trans/cis}}$ were measured in the indicated solvents at 25 °C by integration of ¹³C or ¹⁹F NMR spectra.

try,⁵⁵ but the configuration is retained here because neighboring-group participation of the nitrogen (or possibly the amide oxygen) leads to a double inversion of stereochemistry at the bridge carbon during the fluorination reaction. The stereochemistry of the fluorine on C₁^γ₁ (elsewhere, C-5^{48,31}) was confirmed by analyzing the coupling constants between H₁^γ₁ (H-5) and H₁^γ₂₁ (H-6_{syn}), which is 7.4 Hz. This four-bond W-plan coupling was not observed between the fluorine and either proton on C₁^γ₂ (C-6), nor between H₁^γ₁ and H₁^γ₂₂ (H-6_{anti}), as the latter two sets of nuclei are not in the appropriate arrangement for such coupling.⁵⁶

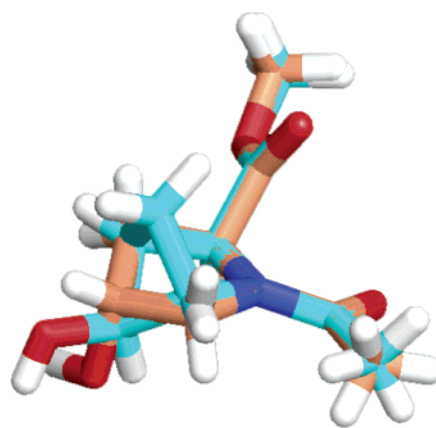
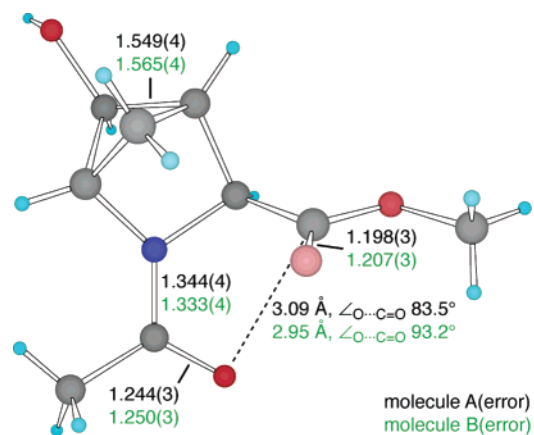
An attractive feature of the methanoproline derivatives is that both pyrrolidine ring puckers are incorporated into a single framework, which allows dissection of the relative contributions of ring pucker and inductive effects on the conformation of substituted prolines. The hydroxyl- and fluoro-substituted methanoproline derivatives are analogues of (2*S*,4*S*)-4-hydroxyproline (hyp) and (2*S*,4*S*)-4-fluoroproline (flp), respectively, and the substituent is in an orientation analogous to the disfavored C^γ-exo pucker.^{40,42} In other words, the bicyclic structure fixes the hydroxyl and fluoro groups of compounds **6** and **7** to be in an antithetical conformation—anti rather than gauche to the pyrrolidine nitrogen about the C^δ₂–C^γ₁ bond.

We measured the amide trans/cis ratios of **5** and **6** by using NMR spectroscopy. We used ¹³C NMR spectroscopy because the ¹H resonances of both methyl groups overlapped with those of other protons from the bicyclic ring system. ¹³C NMR spectra were obtained with ¹H-decoupling enabled only during the acquisition phase of the pulse sequence, allowing for no NOE buildup and thus enabling quantitative integration of the relevant peaks. The trans/cis ratios were measured in CDCl₃, 1,4-dioxane-*d*₈, and D₂O and are listed in Table 2. The trans/cis ratios of **7** were measured by ¹⁹F NMR spectroscopy, as the two ¹⁹F resonances were well-resolved. We found little variation among the three derivatives in a particular solvent. The trans/cis ratios in deuterated 1,4-dioxane and chloroform were all similar, and the ratios in water were somewhat greater than those in the organic solvents.³⁶ These data demonstrate that rigidifying the pyrrolidine ring of proline derivatives by adding a methano bridge abolishes any inductive effect exerted by a fluoro- or a hydroxyl group on the trans/cis ratio of its peptide bond. Apparently, an electronegative substituent on the flexible pyrrolidine ring of proline affects the trans/cis ratio by altering the pucker of the ring.

Next, we determined the crystalline structure of Ac-methano-hyp-OMe (**6**) by X-ray diffraction analysis.

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**FIGURE 3.** Superposition of crystalline structures of Ac-methano-hyp-OMe (**6**, cyan) and Ac-Hyp-OMe (orange).⁵⁷**FIGURE 4.** Ball-and-stick diagram showing bond lengths that differ statistically in the two molecules of the X-ray structure of crystalline Ac-methano-hyp-OMe (**6**). The O₀...C₁=O₁ bond lengths and angles are also shown.

There are two crystallographically independent molecules in the unit cell, as shown in Figure 2. The ring structures of the two molecules are superimposable, while the ester and amide conformations vary slightly relative to one another. The ϕ angles (C_{*i*-1}–N_{*i*}–C_{*i*}–C_{*i*}) differ by 3.1(3)°, the ψ angles (here, N_{*i*}–C_{*i*}–C_{*i*}–O_{*i*+1}) differ by 18.0(4)°, and the ω angles (C_{*i*}^α–C_{*i*}–N_{*i*+1}–C_{*i*+1}^α) differ by 2.6(2)°. A superposition of the structures of Ac-methano-hyp-OMe (**6**) and Ac-Hyp-OMe⁵⁷ is shown in Figure 3 and clearly depicts the antipodal configuration of the hydroxyl groups on C^γ.

All relevant structural parameters support the presence of a stronger n → π* interaction³⁸ in molecule B than in molecule A of crystalline Ac-methano-hyp-OMe (**6**), as shown in Figure 4.⁴⁰ For example, the O₀...C₁ distance is 2.949(4) Å in molecule B, but 3.092(3) Å in molecule A. In addition, the C₁=O₁ bond length is 1.207(3) Å in molecule B, but 1.198(3) Å in molecule A, and the ⁺N₁=C₀–O₀[−] amidic resonance structure appears to be more prevalent in molecule B, which has a shorter N₁–C₀ bond and a longer C₀–O₀ bond than does molecule A. Finally, the O₀...C₁=O₁ angle is closer to the Bürgi–Dunitz optimum of 109°^{58–62} in molecule B [93.2(2)°] than in

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TABLE 3. Values of ϕ and ψ Dihedral Angles for the *trans*-Amide Isomers of Ac-Xaa-OMe^a

Xaa	ring pucker	ϕ (deg)	ψ (deg)	ref
Pro	C γ -exo	-58.6	143.0	40
Pro	C γ -endo	-70.0	152.1	40
Hyp (1)	C γ -exo	-62.0	156.4	57
Hyp (2)	C γ -exo	-50.9	145.2	57
Flp	C γ -exo	-59.2	140.8	40
flp	C γ -endo	-76.4	169.0	40
methano-hyp (1) (6)		-65.0	169.0	this work
methano-hyp (2) (6)		-61.9	153.5	this work

^a Dihedral angles of Xaa = Pro, Flp, and flp are from density functional theory calculations; dihedral angles of Xaa = Hyp and methano-hyp are from X-ray diffraction analysis of crystalline molecules.

molecule A [83.6(2)°]. All of these structural parameters are consistent with greater donation of electron density from the nonbonding electrons of O₀ to the antibonding orbital of the C₁=O₁ bond in molecule B, as expected from a stronger n \rightarrow π^* interaction.⁴⁰ Moreover, the congruence of these five structural parameters (three bond lengths, an atom...atom distance, and an atom...atom-atom angle) provides additional support for the existence of a meaningful n \rightarrow π^* interaction in Ac-methano-hyp-OMe (6) as well as a benchmark for detecting n \rightarrow π^* interactions in other derivatives of proline.

The *trans/cis* ratios for compounds 5–7 in D₂O are intermediate between those of Ac-Pro-OMe and Ac-hyp-OMe or Ac-flp-OMe (Table 1), indicating that the n \rightarrow π^* interactions in methanoproline derivatives are probably weaker than those found in Pro and 4*R*-substituted prolines but stronger than those in the 4*S*-substituted prolines. Likewise, the ϕ and ψ angles of Ac-methano-hyp-OMe (6) are intermediate between those of the endo- and exo-puckers of proline derivatives (Table 3), which is consistent with its intermediate *trans/cis* ratio.

Conclusions

Electronegative substituents in the 4-position of proline residues had been shown to have a substantial effect on the *trans/cis* ratio of their peptide bonds (Table 1).^{36–39} Here, constraining the pucker of the pyrrolidine ring of 4-substituted proline residues with a one-carbon bridge, as in compounds 5–7, was shown to abolish the effect of the electronegative substituents on the *trans/cis* ratio (Table 2). Thus, changes in *trans/cis* ratio arise from changes in ring pucker. This finding suggests that pyrrolidine ring pucker is a key determinant of the stability (or instability) endowed by 4-substituted proline residues on collagen.

Experimental Section

General Procedures. Thin-layer chromatography was performed on precoated plates of silica gel GF 250 μ m. Column chromatography was performed on silica gel, Merck grade 60

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(230–400 mesh). Reagent chemicals were obtained from commercial suppliers, and reagent grade solvents were used without further purification. ¹H NMR spectra were recorded at 300 MHz and ¹³C NMR spectra were recorded at 75 MHz in CDCl₃, unless noted otherwise. Both the ¹H and ¹³C NMR spectra are often complicated by the presence of carbamate conformers and pairs of ¹³C NMR lines due to a single carbon, identified using proton-carbon correlation experiments, have been presented as pairs. Chemical shifts are expressed in ppm relative to internal TMS (¹H) or solvent CDCl₃ (¹³C). High-resolution mass spectra were performed at the University of Pennsylvania, Drexel University, or Merck Research Laboratories (West Point, PA) using FAB ionization methods.

For purposes of nomenclature, 3-*exo* orientation on 2-azabicyclo[2.1.1]hexanes refers to the 3-substituent oriented toward the bridge containing the lower priority attached 5- or 6-substituent. *N*-(Benzyloxycarbonyl)-2-hydroxymethyl-1,2-dihydropyridine and *N*-(benzyloxycarbonyl)-2-hydroxymethyl-2-azabicyclo[2.2.0]hex-5-ene were prepared according to our previously described procedures for *N*-(methoxycarbonyl)-2-hydroxymethyl-1,2-dihydropyridine and *N*-(methoxycarbonyl)-2-hydroxymethyl-2-azabicyclo[2.2.0]hex-5-ene.⁴⁷

***N*-(Benzyloxycarbonyl)-5-*anti*,6-*anti*-dibromo-3-(*p*-nitrophenylsulfonyloxy)methyl-2-azabicyclo[2.1.1]hexane (9).** A solution of bromine (2.56 g, 1.7 equiv) in CH₂Cl₂ (50 mL) was added dropwise to a cold (–5 to 0 °C) solution of nosylate 8 (4.0 g, 9.3 mmol) in CH₂Cl₂ (50 mL), and the resulting solution was stirred for 2 h. The temperature was then raised to 25 °C, and the reaction mixture was stirred for an additional 10 h. The solution was diluted with ether and washed with aqueous sodium bisulfite (10% w/v) until no brown color remained. The organic layer was washed with water, dried over sodium sulfate, and filtered. The solvent was removed in vacuo to provide a crude oil. Purification by flash chromatography (4:1 hexane/EtOAc) afforded 2.74 g (50%) of rearranged dibromide 9 as the major product with *R*_f = 0.47 (1:1 ether/hexane): ¹H NMR (CDCl₃) δ 8.39 (br, 2H), 8.06 (br, 2H), 7.50–7.26 (m, 5H), 5.13 (s, 2H), 4.56 (m, 2H), 4.11 (m, 3H), 3.96 (d, *J* = 7.8 Hz, 1H), 3.19 (d, *J* = 7.2 Hz, 1H); ¹³C NMR δ 155.0, 150.9, 140.9, 135.2, 129.2, 128.7, 128.2, 125.5, 124.6, 68.1, 68.0, 66.1, 59.8, 51.7, 50.9, 46.5; HRMS *m/z* 610.9090, 612.9081, 614.9065, calcd for C₂₀H₁₈N₂O₇S⁷⁹Br⁷⁹BrNa (M + Na) 610.9099, C₂₀H₁₈N₂O₇S⁷⁹Br⁸¹BrNa (M + Na) 612.9079; and C₂₀H₁₈N₂O₇S⁸¹Br⁸¹BrNa (M + Na) 614.9058. In addition, 750 mg (25%) of azatricycle byproduct 10 was also isolated: *R*_f = 0.40 (1:1 ether/hexane); ¹H NMR (CDCl₃) δ 7.35 (m, 5H), 5.16, 5.10 (two d, *J* = 12.6 Hz, 2H), 4.93 (dd, *J* = 2.4, 3.9, 5.4 Hz, 1H), 4.72 (dd, *J* = 3.6, 5.7 Hz, 1H), 4.64 (dd, *J* = 3.9, 10.2 Hz, 1H), 4.42 (dd, *J* = 3.6, 3.6 Hz, 1H), 4.21, 4.11 (s, 1H), 3.92 (ddd, *J* = 3.6, 5.4, 5.7 Hz, 1H), 3.76 (dd, *J* = 2.4, 10.2 Hz, 1H); ¹³C NMR δ 153.4, 135.6, 128.0, 127.7, 127.3, (85.7, 85.5), (73.4, 72.5), (66.5, 66.2), (65.1, 64.4), (63.8, 63.0), (49.0, 48.5), 42.5; HRMS *m/z* 346.0055, calcd for C₁₄H₁₄NO₃Na⁷⁹Br (M + Na), 346.0055; and 348.0036, calcd for C₁₄H₁₄NO₃Na⁸¹Br (M + Na), 348.0034.

***N*-(Benzyloxycarbonyl)-5-*anti*,6-*anti*-dibromo-3-acetoxymethyl-2-azabicyclo[2.1.1]hexane (11).** To the dibromide 9 (300 mg, 0.50 mmol) in dry DMF (10 mL) was added CsOAc (336 mg, 1.75 mmol, 3.5 equiv). The solution was heated to 60 °C for 30 h, cooled, filtered through Celite, and concentrated under reduced pressure. The reaction was diluted with EtOAc and washed with water and brine. The organic layer was dried over sodium sulfate and filtered. Removal of solvent in vacuo and flash chromatography (4:1 hexane/EtOAc) afforded 205.6 mg (92%) of acetate 11: *R*_f = 0.46 (1:1 ether/hexane); ¹H NMR δ 7.41 (s, 5H), 5.22 (m, 2H), 4.62 (d, *J* = 7.3 Hz, 1H), 4.53 (d, *J* = 6.2 Hz, 1H), 4.35 (d, *J* = 7.6 Hz, 1H), 4.16 (b, 2H), 4.05 (d, *J* = 7.6 Hz, 1H), 3.19 (d, *J* = 7.3 Hz, 1H), 2.12 (s, 3H); ¹³C NMR δ 170.4, 155.1, 135.8, 128.7, 128.5, 128.2, 67.9, 66.3, 62.7, 60.4, 52.3, 51.4, 47.3, 20.7; HRMS *m/z* 467.9411, 469.9405, 471.9391, calcd for C₁₆H₁₇NO₄⁷⁹Br⁷⁹BrNa

(M + Na) 467.9422; C₁₆H₁₇NO₄⁷⁹Br⁸¹BrNa (M + Na) 469.9402; and C₁₆H₁₇NO₄⁸¹Br⁸¹BrNa (M + Na) 471.9381.

N-(Benzyloxycarbonyl)-3-acetoxymethyl-2-azabicyclo[2.1.1]hexane (12). Dibromide **11** (474 mg, 1.06 mmol) and AIBN (47.4 mg, 0.29 mmol) were dissolved in benzene (25 mL), and the system was purged with Ar(g) for 10 min. Tributyltin hydride (713 ≥ L, 772 mg, 2.65 mmol) was added via syringe, and the resulting solution was heated at reflux for 2 h. The reaction mixture was cooled to rt, and the solvent was removed in vacuo. The residue was purified by flash chromatography (1:5 ether/hexane) to give 277 mg (91%) of acetate **12**: *R*_f = 0.46 (1:1 ether/hexane); ¹H NMR δ 7.42 (s, 5H), 5.21 (s, 2H), 4.50 (dd, *J* = 4.3, 6.7 Hz, 1H), 4.40 (d, *J* = 7.1 Hz, 1H), 4.14 (m, 1H), 3.96 (b, 1H), 2.83 (d, *J* = 7.1 Hz, 1H), 2.05 (s, 3H), 1.88–1.42 (m, 4H); ¹³C NMR δ 170.7, 156.9, 136.7, 128.5, 128.0, 127.8, 66.8, 64.1, 61.3, 58.2, 41.8, 41.1, 36.4, 20.8; HRMS *m/z* 290.1386, calcd for C₁₆H₂₀NO₄ (M + H) 290.1392.

N-(Benzyloxycarbonyl)-3-hydroxymethyl-2-azabicyclo[2.1.1]hexane (13). To acetate **12** (300 mg, 1.04 mmol) in methanol (25 mL) were added K₂CO₃ (450 mg, 3.0 mmol) and NaHCO₃ (830 mg, 9.0 mmol). The solution was stirred at 25 °C for 2 h and then concentrated in vacuo to remove ca. 90% of the methanol. The resulting slurry was diluted with water and extracted with ether. The combined ether extracts were washed with brine and dried over Na₂SO₄(s). The solvent was removed in vacuo, and the residue was purified by flash chromatography (1:1 ether/hexane) to afford 218 mg (85%) of alcohol **13**: *R*_f = 0.23 (1:2 hexane/ether); ¹H NMR δ 7.41 (s, 5H), 5.22 (s, 2H), 4.59 (b, 1H), 4.45 (d, *J* = 7.18 Hz, 1H), 3.96–3.79 (br, 2H), 2.75 (b, 1H), 2.02–1.50 (m, 4H); ¹³C NMR δ 157.9, 136.4, 128.5, 128.1, 127.9, 67.3, 66.0, 62.8, 61.4, 42.6, 41.2, 37.7; HRMS *m/z* 270.1113, calcd for C₁₄H₁₇NO₃Na (M + Na) 270.1106. Alcohol **13** was admixed with about 10% of a second inseparable alcohol in which the *N*-benzyloxycarbonyl group was exchanged for *N*-methoxycarbonyl as shown by a small peak at δ 3.80 (s). This impurity was removed at the next stage.

N-(Benzyloxycarbonyl)-3-methoxycarbonyl-2-azabicyclo[2.1.1]hexane (14). To a solution of the slightly impure alcohol **13** (600 mg, 2.45 mmol) in CH₂Cl₂ (7 mL) containing TEMPO (5 mg, 0.032 mmol) was added a solution of saturated aqueous NaHCO₃ (4.8 mL) containing KBr (26.4 mg, 0.22 mmol) and tetrabutylammonium chloride (35.2 mg, 0.13 mmol). The mixture was cooled to 0 °C, and a solution of NaOCl (4–6% w/v, 12 mL), saturated aqueous NaHCO₃ (2.64 mL), and brine (5.3 mL) was added dropwise over 45 min. The two layers were separated, and the organic layer was washed with water. The combined aqueous extracts were acidified with 4 M HCl and extracted extensively with ethyl acetate. The organic layer was dried over Na₂SO₄(s) and concentrated in vacuo. The crude acid was dissolved in isopropyl alcohol (15 mL) and hexane (15 mL), and trimethylsilyldiazomethane (2.0 M, 1.225 mL, 2.45 mmol) was added. The mixture was stirred at 25 °C for 30 min, and then the solvent was removed in vacuo. The residue was purified by flash chromatography, eluting with 4:1 hexane/ether to afford 539 mg (80%) of ester **14**: *R*_f = 0.40 (1:1 hexane/ether); ¹H NMR δ 7.37 (s, 5H), 5.22 (m, 2H), 4.53 (br, 1H), 4.34 (br, 1H), 3.79 (br, 3H), 3.05 (br, 1H), 2.11–1.48 (m, 4H); ¹³C NMR δ 171.4, 156.8, 66.9, 60.9, 60.2, 52.2, 43.4, 42.9, 37.7; HRMS *m/z* 298.1070, calcd for C₁₅H₁₇NO₄Na (M + Na) 298.1055. This ester contained 40 mg of a minor byproduct, the *N*-(methoxycarbonyl)-3-ester: *R*_f = 0.40 (1:1 hexane/ether); ¹H NMR δ 4.40 (br, 1H), 4.22 (br, 1H), 3.73 (s, 3H), 3.67 (s, 3H), 2.97 (m, 1H), 2.03–1.39 (m, 4H); ¹³C NMR δ 171.3, 156.2, 61.0, 60.1, 52.4, 52.1, 43.3, 42.7, 37.2; HRMS *m/z* 200.0918, calcd for C₉H₁₄NO₄ (M + H) 200.0923. A sample of the major product, ester **14** (25 mg), was separated from the mixture (60 mg) by collection of the first fraction upon chromatography (1:10 ether/hexane); the remainder (20 mg) was a mixture of esters. A pure sample of the minor ester was obtained following *N*-debenzylation of the major isomer (below).

N-¹³CH₃-Labeled-acetyl-3-methoxycarbonyl-2-azabicyclo[2.1.1]hexane (5). To a solution of ester **14** (200 mg, 0.73 mmol) in methanol (20 mL) was added Pd/C (100 mg). The mixture was stirred at 25 °C under a H₂(g)-filled balloon for 2 h and then filtered through Celite. The solvent was removed in vacuo to afford 105 mg of crude amine: *R*_f = 0.60 (5:1 EtOAc/methanol); ¹H NMR (CDCl₃) δ 4.70 (br, 1H), 4.32 (d, *J* = 7.2 Hz, 1H), 3.84 (s, 3H), 3.18 (d, *J* = 7.2 Hz, 1H), 2.20 (d, *J* = 7.2 Hz, 1H), 2.16 (d, *J* = 7.2 Hz, 1H), 2.00 (dd, *J* = 10.6, 7.2 Hz, 1H), 1.63 (dd, *J* = 10.6, 7.2 Hz, 1H); ¹³C NMR (CDCl₃) δ 171.1, 60.4, 59.0, 52.2, 43.9, 42.2, 37.0; HRMS *m/z* 142.0873, calcd for C₇H₁₂NO₂ (M + H) 142.0868. To a solution of the amine in dry CH₂Cl₂ (10 mL) at 0 °C was added (dimethylamino)pyridine (DMAP, 759 mg, 6.21 mmol), followed by dropwise addition of ¹³CH₃COCl (170.8 mg, 2.15 mmol). The mixture was stirred for an additional 30 min at 0 °C, allowed to warm slowly to 25 °C, and then stirred for 2 h. Water was added, and the aqueous layer was extracted with EtOAc. The combined organic layers were dried over Na₂SO₄(s) and filtered. The solvent was removed in vacuo, and the residue was purified by flash chromatography eluting with EtOAc to give 107 mg (80%) of **5** as a colorless oil: *R*_f = 0.50 (15:1 EtOAc/MeOH); ¹H NMR δ 4.8–4.4 (m, 1H, two conformers), 4.34 (m, 1H), 3.80 (m, 3H), 3.06 (m, 1H), 2.36–1.40 (m, 7H); ¹³C NMR δ (170.8, 169.4), 168.3 (d, *J* = 51.6 Hz), (62.5, 61.0), (59.6, 59.0), (52.5, 52.1), (44.0, 42.9), (42.6, 42.3), (37.8, 36.5) (21.6, 21.5, labeled ¹³C); HRMS *m/z* 185.1002, calcd for C₈¹³CH₁₄NO₃ (M + H) 185.1007.

N-(Benzyloxycarbonyl)-5-anti-hydroxy-6-anti-bromo-3-endo-(*p*-nitrophenylsulfonyloxy)methyl-2-azabicyclo[2.1.1]hexane (15). To the nosylate **8** (2.00 g, 4.65 mmol) in THF (16 mL) and H₂O (8 mL) at 0 °C was added *N*-bromosuccinimide (2.51 g, 13.9 mmol) in small portions so that the temperature never exceeded 0 °C. Upon completion of addition, the solution was warmed to rt and stirred for 2.5 h, diluted with water, and extracted extensively with chloroform. The combined organic layers were washed with brine and dried over Na₂SO₄(s). The solvent was removed in vacuo to give an oil, which was purified by flash chromatography (3:1 ether/hexane) to provide 1.71 g (70%) of bromohydrin **15**: *R*_f = 0.25 (3:1 ether/hexane); ¹H NMR δ 8.43 (br, 2H), 8.14 (br, 2H), 7.47–7.34 (m, 5H), 5.15 (s, 2H), 4.55 (br, 1H), 4.44 (d, *J* = 7.5 Hz, 1H), 4.22 (m, 2H), 4.10 (m, 2H), 3.43 (br, 1H), 3.08 (d, *J* = 7.5 Hz, 1H); ¹³C NMR δ 155.1, 150.8, 141.0, 135.4, 129.2, 128.7, 128.5, 128.1, 124.6, 85.9, 68.2, 67.8, 65.8, 58.3, 51.8, 47.7; HRMS *m/z* 548.9953, calcd for C₂₀H₁₉N₂O₈NaS⁷⁹Br (M + Na) 548.9943; *m/z* 550.9929, calcd for C₂₀H₁₉N₂O₈NaS⁸¹Br (M + Na) 550.9923.

N-(Benzyloxycarbonyl)-5-anti-(*tert*-butyldimethylsilyloxy)-6-anti-bromo-3-endo-(*p*-nitrophenylsulfonyloxy)-methyl-2-azabicyclo[2.1.1]hexane (16). To a solution of nosylate **15** (2.33 g, 4.4 mmol) in DMF (5 mL) were added *tert*-butyldimethylsilyl chloride (TBDMSCl) (0.80 g, 5.3 mmol) and imidazole (0.76 g, 11 mmol). The resulting solution was stirred at rt until no starting material remained (21 h), as determined by TLC, and then diluted with diethyl ether (20 mL) and water (10 mL). The two layers were separated, and the aqueous layer was extracted with diethyl ether. The organic layer was washed with brine, dried over Na₂SO₄(s), and filtered. The solvent was removed in vacuo to give the crude product, which was purified by flash chromatography to provide 1.76 g (62%) of the desired *O*-silyl ether **16** as an oily white solid: *R*_f = 0.64 (2:1 ether/hexane); ¹H NMR δ 8.42 (br, 2H), 8.13 (br, 2H), 7.40–7.31 (5H), 5.13 (m, 2H), 4.53 (m, 1H), 4.28 (d, *J* = 7.5 Hz, 1H), 4.20 (d, *J* = 7.8 Hz, 1H), 4.15 (d, *J* = 9.9 Hz, 1H), 4.12 (d (buried), *J* = 7.8 Hz, 1H), 4.06 (m, 1H), 2.90 (d, *J* = 7.2 Hz, 1H); ¹³C NMR δ 157.7, 150.9, 140.9, 129.3, 124.6, 135.6, 128.7, 128.5, 128.1, 84.7, 68.4, 67.7, 66.2, 58.1, 51.6, 45.7, 25.6, 17.9, –5.04; HRMS *m/z* 663.0792, calcd for C₂₆H₃₃N₂O₈NaSi⁷⁹Br (M + Na), 663.0808; *m/z* 665.0773, calcd for C₂₆H₃₃N₂O₈NaSi⁸¹Br (M + Na) 665.0788.

N-Benzyloxycarbonyl-5-anti-(tert-butyl)dimethylsilyloxy-6-anti-bromo-3-endo-acetoxymethyl-2-azabicyclo[2.1.1]hexane (17). To a solution of *O*-silyl ether **16** (128 mg, 0.20 mmol) in toluene (30 mL) were added cesium acetate (200 mg, 1.0 mmol, 5 equiv), DMAP (24.5 mg, 0.20 mmol), and 18-crown-6 (7 mg). The resulting solution was heated at reflux for 8 h and then cooled and filtered through Celite. The solvent was concentrated in vacuo, and the resulting slurry was diluted with EtOAc and then washed with water followed by brine. The organic layer was dried over Na₂SO₄(s) and filtered. Removal of solvent in vacuo and flash chromatography afforded 72 mg (72%) of acetate **17** as a yellow oil: *R*_f = 0.32 (1:2 ether/hexane); ¹H NMR δ 7.35 (s, 5H), 5.19 (dd, *J* = 7.5 Hz, 2H), 4.43 (dd, *J* = 3.6, 10.8 Hz, 1H), 4.28 (d, *J* = 7.2 Hz, 1H), 4.26 (d, *J* = 7.2 Hz, 1H), 4.14 (d, *J* = 7.2 Hz, 1H), 4.05 (br, 2H), 2.82 (d, *J* = 7.2 Hz, 1H), 2.06 (s, 3H), 0.91 (s, 9H), 0.07 (s, 6H); ¹³C NMR δ 170.4, 155.0, 135.9, 128.5, 128.2, 128.0, 84.8, 67.4, 66.1, 63.0, 58.5, 52.1, 46.5, 25.7, 20.7, 17.9, -5.1; HRMS *m/z* 520.2243, calcd for C₂₂H₃₂NO₅NaSi⁷⁹Br (M + Na) 520.1131; *m/z* 522.1115, calcd for C₂₂H₃₂NO₅NaSi⁸¹Br (M + Na) 522.1110.

N-Benzyloxycarbonyl-5-anti-(tert-butyl)dimethylsilyloxy-3-exo-acetoxymethyl-2-azabicyclo[2.1.1]hexane (18). To a solution of acetate **17** (670 mg, 1.3 mmol) in benzene (50 mL) were added tributyltin hydride (1.18 g, 4.1 mmol) and AIBN (22 mg, 0.13 mmol). The resulting mixture was heated at reflux for 3 h. The solvent was removed in vacuo, and the crude product was purified by flash chromatography to give 540 mg (94%) of acetate **18** as a colorless oil: *R*_f = 0.53 (1:1 ether/hexane); ¹H NMR δ 7.40 (s, 5H), 5.20 (dd, *J* = 12.3 Hz, 2H), 4.44 (dd, *J* = 11.1, 4.2 Hz, 1H), 4.18 (d, *J* = 7.2 Hz, 1H), 4.17 (m, 1H), 4.11 (br, 1H), 3.97 (d, *J* = 7.2 Hz, 1H), 2.86 (br d, *J* = 8.1 Hz, 1H), 2.62 (dd, *J* = 7.2, 3.0 Hz, 1H), 2.09 (s, 3H), 1.84 (dd, *J* = 8.1, 7.2 Hz, 1H), 0.93 (s, 9H), 0.10 (s, 6H); ¹³C NMR δ 170.6, 156.5, 136.5, 128.4, 128.0, 127.8, 82.4, 66.9, 64.2, 63.4, 56.8, 47.4, 32.8, 25.7, 20.8, 17.9, -4.96; HRMS *m/z* 442.2041, calcd for C₂₂H₃₃N₅NaSi (M + Na) 442.2026.

N-Benzyloxycarbonyl-5-anti-tert-butyl)dimethylsilyloxy-3-exo-hydroxymethyl-2-azabicyclo[2.1.1]hexane (19). To a solution of acetate **18** (302 mg, 0.72 mmol) in methanol (10 mL) were added potassium carbonate (357 mg, 2.2 mmol) and sodium bicarbonate (544 mg, 6.5 mmol). The resulting mixture was stirred at rt for 1 h. The bulk of the solvent was removed in vacuo, and the resulting mixture was diluted with water and extracted with EtOAc. The combined organic extracts were washed with brine, dried over Na₂SO₄(s), and concentrated in vacuo to give crude product (240 mg). Flash chromatography provided 207 mg (76%) of alcohol **19** as a colorless liquid: *R*_f = 0.30 (1:1 ether/hexane); ¹H NMR δ 7.40 (m, 5H), 5.21 (m, 2H), 4.34 (br, 1H), 4.21 (dd, *J* = 1.5, 7.5 Hz, 1H), 4.03 (d, 1H, *J* = 7.2 Hz, 1H), 3.99 (br, 1H), 3.80 (br, 2H), 2.84 (dt, *J* = 2.7, 8.1, 1.5 Hz, 1H), 2.55 (m, 1H), 1.79 (dd, *J* = 8.1, 7.2 Hz, 1H), 0.94 (s, 9H), 0.06 (s, 6H); ¹³C NMR δ 157.7, 136.3, 128.5, 128.2, 127.9, 82.9, 67.3, 65.4, 64.5, 61.7, 47.3, 34.0, 25.7, 18.0, -4.9; HRMS *m/z* 400.1930, calcd for C₂₀H₃₁NO₄NaSi (M + Na) 400.1920.

N-Benzyloxycarbonyl-5-anti-tert-butyl)dimethylsilyloxy-3-exo-carboxy-2-azabicyclo[2.1.1]hexane (20). To a solution of alcohol **19** (39 mg, 0.085 mmol) in dichloromethane (0.5 mL) containing TEMPO (0.1 mg) was added a solution of saturated NaHCO₃ (0.2 mL) containing KBr (0.9 mg) and tetrabutylammonium chloride (1.2 mg). The mixture was cooled to 0 °C, and a solution of NaOCl (0.21 mL), saturated NaHCO₃(aq) (0.1 mL), and saturated NaCl(aq) (0.18 mL) was added dropwise over 45 min. The two layers were separated, and the organic layer was extracted with water. The aqueous extracts were combined and acidified with aqueous HCl (10% w/v), and the resulting solution was extracted with EtOAc. The combined organic extracts were dried over Na₂SO₄(s). The solvent was removed to give 37 mg (93%) of the desired carboxylic acid **20**: ¹H NMR (75 °C) δ 10.48 (br, 1H), 7.39 (s, 5H), 5.25 (dd, *J* = 12.3 Hz, 2H), 4.37 (s, 1H), 4.26 (dd, *J* = 7.5,

1.2 Hz, 1H), 4.06 (dd, *J* = 7.2 Hz, 1H), 2.97 (d, br, *J* = 8.1 Hz, 1H), 2.94 (d, *J* = 7.5 Hz, 1H), 2.15 (dd, *J* = 7.2, 8.1 Hz, 1H), 0.96 (s, 9H), -0.13 (s, 6H); ¹³C NMR δ 172.9, 156.9, 135.8, 128.4, 128.1, 127.8, 82.2, 67.3, 64.1, 59.4, 48.0, 34.5, 25.5, 17.9, -4.8; HRMS *m/z* 392.1894, calcd for C₂₀H₃₀NO₅Si (M + H) 392.1893.

N-Benzyloxycarbonyl-5-anti-tert-butyl)dimethylsilyloxy-3-exo-methoxycarbonyl-2-azabicyclo[2.1.1]hexane (21). To a solution of acid **20** (75 mg, 0.19 mmol) in hexane (1.5 mL) and 2-propanol (0.75 mL) was added a 2 M solution of trimethylsilyldiazomethane in hexane (383 μL, 0.766 mmol). The resulting mixture was stirred under argon for 0.5 h. The solvent was removed in vacuo to give 80 mg (100%) of ester **21**: *R*_f = 0.35 (1:2 ether/hexane); ¹H NMR (75 °C) δ 7.38 (s, 5H), 5.27 (d, *J* = 12.3 Hz, 1H), 5.16 (d, *J* = 12.3 Hz, 1H), 4.33 (s, 1H), 4.26 (dd, *J* = 7.5, 1.5 Hz, 1H), 4.05 (d, *J* = 7.2 Hz, 1H), 3.76 (s, 3H), 2.90 (ddd, *J* = 8.1, 3.3, 1.5 Hz, 1H), 2.82 (dd, *J* = 7.5, 3.3 Hz, 1H), 2.04 (dd, *J* = 8.1, 7.2 Hz, 1H), 0.97 (s, 9H), 0.1 (s, 6H); ¹³C NMR δ 170.5, 156.7, 136.5, 128.4, 128.0, 127.7, 82.7, 67.0, (64.3 and 63.5), 58.7, 52.3, (49.1 and 48.8), (33.7, 33.2), 25.6, 18.0, -4.9; HRMS *m/z* 406.2046, calcd for C₂₁H₃₂NO₅Si (M + H) 406.2049.

5-anti-tert-Butyl)dimethylsilyloxy-3-exo-methoxycarbonyl-2-azabicyclo[2.1.1]hexane (22). To a solution of ester **21** (810 mg, 2.0 mmol) in methanol (40 mL) was added 10% Pd/C (200 mg). The solution was stirred at 25 °C under a H₂(g)-filled balloon for 2 h and then filtered through Celite. The solvent was removed in vacuo to give an oil. Purification by flash chromatography (1:1 hexane/EtOAc) afforded 483 mg (85%) of the desired amine **22**: ¹H NMR δ 4.13 (m, 1H), 3.79 (m, 4H), 3.44 (m, 1H), 2.79 (m, 1H), 2.69 (m, 1H), 2.22 (br, 1H), 1.45 (m, 1H), 0.94 (s, 9H), 0.11 (s, 6H); ¹³C NMR δ 174.1, 82.8, 62.8, 58.4, 52.3, 49.7, 33.0, (25.7 and 22.6), -4.9; HRMS *m/z* 272.1672, calcd for C₁₃H₂₆NO₃Si (M + H) 272.1682.

N-(2-¹³CH₃-Acetyl)-5-anti-tert-butyl)dimethylsilyloxy-3-exo-methoxycarbonyl-2-azabicyclo[2.1.1]hexane (23). To a solution of amine **22** (483 mg, 1.6 mmol) in dry methylene chloride (20 mL) at 0 °C was added DMAP (1.66 g, 13.6 mmol, 3 equiv), and then ¹³CH₃COCl (372.7 mg, 4.69 mmol) was added dropwise, and the mixture was stirred for 30 min at 0 °C. The reaction mixture was then slowly warmed to 25 °C and stirred for an additional 2 h. Water (5 mL) was added to form two layers, and the water layer was extracted with EtOAc. The combined organic extracts were dried over Na₂SO₄(s) and filtered. The solvent was removed in vacuo, and the residue was purified by flash chromatography to afford 484 g (93%) of amide **23**: *R*_f = 0.20 (1:1 hexane/EtOAc); ¹H NMR δ 4.57–4.24 (m, 1H), 3.98 (m, 2H), 3.71 (m, 3H), 2.84–2.75 (m, 2H), 2.04 and 1.93 (two d, *J* = 128.1 Hz, 3H), 2.06 (t, *J* = 7.5 Hz, 1H), 0.86 (s, 9H), 0.04 (s, 6H); ¹³C NMR (two conformations) δ 170.1 and 169.6, 169.3 (d, *J* = 51.5 Hz) and 168.1 (d, *J* = 51.6 Hz), 82.4 and 82.0, 65.6 and 62.6, 59.5 and 57.6, 52.5 and 52.2, 49.3 and 48.3, 34.0 and 32.7, 25.5, (21.2 and 21.0, labeled methyl), 17.9, -4.1; HRMS *m/z* 315.1830, calcd for C₁₄¹³CH₂₈NO₄Si (M + H) 315.1821.

N-(2-¹³CH₃-Acetyl)-5-anti-hydroxy-3-exo-methoxycarbonyl-2-azabicyclo[2.1.1]hexane (6). To a solution of silyl ether **23** (486.4 mg, 1.48 mmol) in THF (10 mL) at 0 °C was added tetrabutylammonium fluoride monohydrate (TBAF·H₂O, 1.16 g, 4.44 mmol, 3 equiv). The reaction mixture was stirred at 0 °C for 5 min, and then warmed slowly to rt and stirred for an additional 30 min. Removal of solvent in vacuo and flash chromatography afforded 181 mg (90%) of alcohol **6** as a white solid: *R*_f = 0.60 (5:1 EtOAc/MeOH); ¹H NMR (500 MHz) δ (4.73, 4.68) (br, 1H), (4.55–4.53, 4.11–4.09) (m, 1H), (4.31, 4.28) (bs, 1H), (4.04, 3.98) (m, 1H), (3.78, 3.72) (s, 3H), 2.91–2.81 (m, 2H), 2.11 (m, 1H), (2.06, 1.94) (d, *J* = 128.3 Hz, 3H); ¹³C NMR (125 MHz, two conformations) δ 170.0 (d, *J* = 50.3 Hz), 169.9, 168.4 (d, *J* = 51.7 Hz), 81.8, 81.3, 65.3, 62.5, 59.9, 57.9, 52.6, 52.3, 48.5, 47.4, 34.1, 32.8, 21.8, 21.65, (21.58, 21.4 labeled ¹³C); HRMS *m/z* 201.0961, calcd for C₈¹³CH₁₄NO₄ (M + H) 201.0956.

***N*-Acetyl-5-*anti*-fluoro-3-*exo*-methoxycarbonyl-2-azabicyclo[2.1.1]hexane (7).** Bis(2-methoxyethyl)aminosulfur trifluoride (166 mg, 0.75 mmol) was added dropwise via syringe to a solution of *N*-acetyl-5-*anti*-hydroxy-3-*exo*-methoxycarbonyl-2-azabicyclo[2.1.1]hexane **6** (60 mg, 0.3 mmol) in dry CH₂Cl₂ (12 mL) under Ar(g) at -78 °C. The mixture was stirred for 2 h at rt and then heated at reflux for 8 h. The reaction was quenched with water, and the aqueous layer was extracted with CH₂Cl₂. The organic extracts were combined and washed with water and brine, dried over Na₂SO₄(s), and filtered. Removal of solvent in vacuo and flash chromatography (EtOAc) gave 38 mg (63%) of fluoride **7** as an oil: $R_f = 0.55$ (15:1 EtOAc/MeOH); ¹H NMR δ [4.77 (dd, ² $J_{H,F} = 61.6$ Hz, $J = 7.4$ Hz), 4.62 (dd, ² $J_{H,F} = 61.0$ Hz, $J = 7.4$ Hz), 1H], 4.33 (s, 1H), 4.24 (dd, $J = 7.2$, 1.5 Hz, 1H), 3.74 and 3.70 (two s, 3H), 3.01 (m, 1H), 2.79 (m, 1H), 2.25–2.22 (m, 1H), 2.11 and 1.99 (two s, 3H); ¹³C NMR (100 MHz, two conformations) δ 169.6, 169.4, 169.2, 168.1, 97.9 (d, $J_{C,F} = 219.6$ Hz), 97.6 (d, $J_{C,F} = 218.7$ Hz), 63.5 (d, $J_{C,F} = 20.5$ Hz), 60.5 (d, $J_{C,F} = 20.8$ Hz), 58.6, 56.5, 52.8, 52.5, 47.6 (d, $J_{C,F} = 19.2$ Hz), 46.7 (d, $J_{C,F} = 18.7$ Hz), 34.2, 32.8, 30.9, 21.6, 21.6; HRMS m/z 202.0885, calcd for C₉H₁₃NO₃F (M + H) 202.0879.

Measurement of $K_{trans/cis}$ Values of 5–7. Each compound (10–20 mg) was dissolved in CDCl₃ (approximately 1 mL) and the ¹³C NMR (**5** and **6**) or ¹⁹F NMR (**7**) spectrum recorded. The relaxation delay for the measurement of the spectra of **5** and **6** was 10–18 s to allow for full relaxation of the ¹³C nuclei. The spectral baselines were corrected and peaks corresponding to the labeled carbon or the fluorine were integrated with the software package NUTS.⁶³ The samples were then concentrated under reduced pressure and placed under high vacuum overnight to ensure removal of all residual CDCl₃. The resulting samples were dissolved in 1,4-dioxane-*d*₈ (800 μ L), and the spectra were recorded again. The samples were concentrated under reduced pressure and placed under high vacuum overnight. D₂O (800 μ L) was then added to each sample followed by enough CD₃OD to effect full dissolution of the sample. The amount of added CD₃OD was less than 20% of the total volume in each case. The samples were filtered, their spectra were recorded, and the *trans/cis* ratios were determined by integration of the respective resonances.

Crystallization of Ac-methano-hyp-OMe (6). Racemic Ac-methano-hyp-OMe (**6**, 20–30 mg) was dissolved in dichloromethane, and the resulting solution was aliquotted into five vials. A cosolvent (10–20 drops) was added to each vial with a Pasteur pipet: vial 1, hexanes; vial 2, diethyl ether; vial 3, 1,4-dioxane; vial 4, no cosolvent; vial 5, ethyl acetate. The vials were capped loosely and allowed to sit at rt for approximately 2 days. Vial 5 contained the crystals most suitable for X-ray crystallography, and these crystals were used for X-ray diffraction analysis.

X-ray Diffraction Data Collection. An air-stable crystal of Ac-methano-hyp-OMe (**6**) with approximate dimensions 0.50 \times 0.40 \times 0.40 mm³ was selected under oil at ambient conditions and attached to the tip of a glass capillary. The crystal was mounted in a stream of cold nitrogen at 173(2) K and centered in the X-ray beam by using a microscope.

Crystal evaluation and data collection were performed on a Bruker P4/CCD-1000 diffractometer with Mo K α ($\lambda = 0.710$ 73 Å) radiation with a diffractometer-to-crystal distance of 4.999 cm.

Initial cell constants were obtained from three series of ω scans at different starting angles. Each series consisted of 20 frames collected at intervals of 0.3° in a 6° range about ω with an exposure time of 10 s per frame. A total of 69 reflections

were obtained. The reflections were indexed successfully by an automated indexing routine built in the SMART program.⁶⁵ The final cell constants were calculated from a set of 4952 strong reflections from the actual data collection.

Data were collected by using the multirun data collection routine. The reciprocal space was surveyed to the extent of a full sphere to a resolution of 0.80 Å. A total of 12437 data were harvested by collecting one set of 1250 frames with 0.3° scans in ϕ and four sets of 100 frames with 0.3° scans in ω with an exposure time 30 s per frame. This highly redundant data set was corrected for Lorentz and polarization effects. The absorption correction was based on fitting a function to the empirical transmission surface as sampled by multiple equivalent measurements.⁶⁴

Structure Solution and Refinement. The systematic absences in the diffraction data were consistent for the space groups $P1$ and $P\bar{1}$.⁶⁵ The E -statistics strongly suggested the centrosymmetric space group $P\bar{1}$ that yielded chemically reasonable and computationally stable results of refinement.

A successful solution by direct methods provided most non-hydrogen atoms from the E -map. The remaining non-hydrogen atoms were located in an alternating series of least-squares cycles and difference Fourier maps. All non-hydrogen atoms were refined with anisotropic displacement coefficients. All hydrogen atoms were included in the structure factor calculation at idealized positions and were allowed to ride on the neighboring atoms with relative isotropic displacement coefficients. There were two chemically equivalent but crystallographically independent molecules of compound **6** in the asymmetric unit. Because compound **6** crystallized in a centrosymmetric space group, the crystal structure was a racemic mixture of stereoisomers. Several likely intermolecular hydrogen-bonding interactions were observed in the lattice, and formed a series of one-dimensional chains in the ab plane.

The final least-squares refinement of 259 parameters against 3638 data resulted in residuals R (based on F^2 for $I \geq 2\sigma$) and wR (based on F^2 for all data) of 0.0735 and 0.2199, respectively. The final difference Fourier map was featureless.

Acknowledgment. We are grateful to Drs. C. W. Roth, III, and J. A. Hodges for helpful discussions. This work was supported by Grant Nos. AR44276 (NIH to R.T.R.) and CHE-0111208 (NSF to G.R.K.) and by the donors of the Petroleum Research Fund, administered by the American Chemical Society (to G.R.K.). C.L.J. was supported by Chemistry–Biology Interface Training Grant No. GM08506 (NIH). NMR spectra for $K_{trans/cis}$ determinations were obtained at the Magnetic Resonance Facility in the Department of Chemistry at the University of Wisconsin–Madison, which was supported by Grant Nos. CHE-8813550, CHE-9629688, and CHE-9208463 (NSF) and S10 RR08389-01 and S10 RR04981-01 (NIH). The illustration on the cover of this issue was created by H. Adam Steinberg (The Media Lab, Department of Biochemistry, University of Wisconsin–Madison).

Supporting Information Available: ¹H and ¹³C NMR spectra of compounds **5–23** and data from the X-ray diffraction analysis of Ac-methano-hyp-OMe (**6**). This material is available free of charge via the Internet at <http://pubs.acs.org>.

JO049242Y

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(65) All software and sources of the scattering factors are contained in the SHELXTL (version 5.1) program library: Sheldrick, G., Ed. Bruker Analytical X-ray Systems, Madison, WI.

(63) NUTS–NMR Utility Transform Software, Acorn NMR Inc., 7670 Las Positas Road, Livermore, CA 94551.